

The Reaction of Three Sesquiterpene Ethers with *m*-Chloroperbenzoic Acid

Motoo TORI, Masakazu SONO, and Yoshinori ASAKAWA*

Faculty of Pharmaceutical Sciences, Tokushima Bunri University, Yamashiro-cho, Tokushima 770

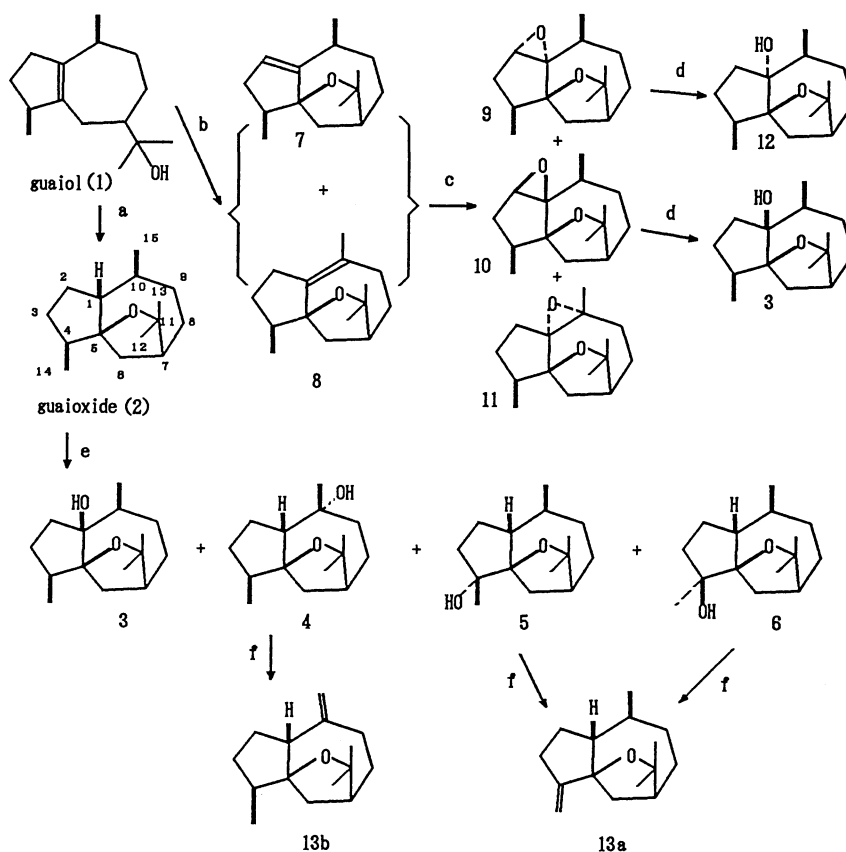
(Received January 20, 1990)

Three sesquiterpene ethers, guaiooxide, a guaiane-type sesquiterpene ether (LB), and maaliioxide have been subjected to reaction with *m*-chloroperbenzoic acid in chloroform under reflux. Guaiooxide afforded four tertiary alcohols, while other two gave secondary alcohols. Structures of the products were determined by spectral data and chemical correlations.

Oxidation reactions at unactivated carbon atoms are difficult but very interesting and important. The known methods include microbial oxidation,¹⁾ mammalian metabolism,²⁾ Barton reaction,³⁾ remote oxidation,⁴⁾ and dry ozonization.⁵⁾ In the previous papers, we reported the oxidation of natural products by *m*-chloroperbenzoic acid (*m*CPBA) yielding hydroxyl or carbonyl compounds as a result of functionalization of unactivated carbon atoms. Application of this method to mono-,⁶⁾ sesqui-,⁷⁾ and triterpenes,⁸⁾ and acyclic compounds⁹⁾ have been published and proved to be a convenient method for introduction of a hydroxyl group. Namely the method is very simple, safe, and applicable to a variety of compounds.

Guaiooxide (**2**) is a tricyclic sesquiterpene ether isolated from guaiac wood oil and its microbial transfor-

mation has been reported.¹⁰⁾ Oxidation reactions by dry ozonization and by the metabolism using rabbits have been studied.¹¹⁾ Radical brominations¹²⁾ have also been investigated. A guaiane-type sesquiterpene ether (a stereoisomer of **2**), LB (**14**), was isolated from the roots of a *Ligularia* genus by T. Takahashi and his group and its structure was determined by degradation.¹³⁾ Radical bromination of LB (**14**) using NBS has been studied by Hirota et al. as a method for functionalization of unactivated carbon atoms.¹³⁾ In connection with our recent study on oxidation reaction of natural and unnatural substances, three sesquiterpene ethers, guaiooxide (**2**), LB (**14**), and maaliioxide (**22**),¹⁴⁾ were subjected to *m*CPBA oxidation. We now describe the details of these reactions.



Scheme 1. a) H^+ ; b) $Pb(OAc)_4$; c) *m*CPBA; d) $LiAlH_4$; e) *m*CPBA/ $CHCl_3$ /reflux; f) $POCl_3$ /Py.

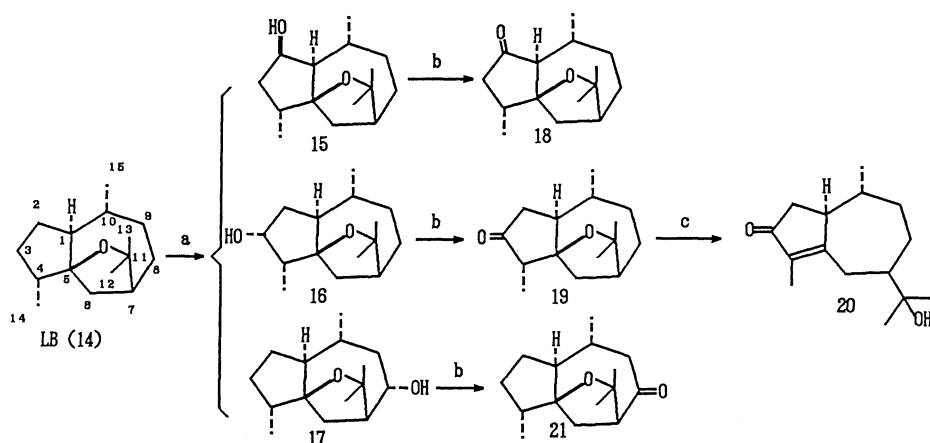
Results and Discussion

Oxidation of Guaioxide (2). Guaioxide (2) was treated with *m*CPBA in chloroform under reflux for 13 h to afford compounds 3, 4, and 5 after column chromatography. Careful purification by HPLC (Finepak Sil) yielded the fourth compound 6. These four compounds were all tertiary alcohols (no proton attached to a carbon bearing the hydroxyl group was observed in the NMR spectra). Compound 3 had two secondary methyl groups, while the others had only one secondary methyl group. These facts suggest that compound 3 has a hydroxyl group at either C-1 or C-7 and others have at either C-4 or C-10, apart from their stereochemistry.

Since 7 α -hydroxyguaioxide is known,¹⁰ compound 3 is inferred to be 1-hydroxyguaioxide. To establish the stereochemistry at 1-position, both 1 α - and 1 β -hydroxyguaioxide, (12) and (3), were prepared from guaiol (1). Guaiol (1) was oxidized by lead tetraacetate according to the literature¹⁵ to afford Δ^1 -dehydroguaioxide (7) in addition to $\Delta^{1(10)}$ -dehydroguaioxide (8), whose formation had not been reported so far.¹⁵ The mixture was treated with *m*CPBA to afford three epoxides 9–11, which were separated by silica-gel column chromatography. It was not easy to assign the stereochemistry of the epoxides 9 and 10 by the NMR spectra. However, the major epoxide 9 and the minor epoxide 10 were suggested tentatively to be 1 α ,2 α -epoxyguaioxide and 1 β ,2 β -epoxyguaioxide, respectively, since the β -face was sterically hindered. The fully substituted epoxide (11) was thus assigned to be 1 α ,10 α -epoxyguaioxide by the same discussion. Each epoxide, 9 and 10, was treated with LiAlH₄ to give the corresponding tertiary alcohol 12 and 3, respectively. As expected, the yield of 12 from 9 was very low due to steric hindrance of the β -face, while that of 3 from 10 was very high. The spectral data of the oxidation product 3 were identical with those of the product, 1 β -hydroxyguaioxide (3), synthesized in an alternative route.

Among the other three products, compound 5 showed the identical spectral data with those reported by microbial oxidation¹⁰ and by dry ozonization.¹¹ Thus, the structure of 5 was determined to be 4 α -hydroxyguaioxide. Since the extensive high-field NMR analysis of the other two alcohols, 4 and 6, did not give the conclusive results, three alcohols, 4, 5, and 6, were subjected to dehydration. The known alcohol 5 gave mainly the exo-olefin 13a on dehydration with POCl₃ at 0°C. Under the same reaction conditions, alcohol 4 afforded the other exo-olefin 13b, while 6 yielded 13a. Thus, the structures of 6 and 4 were assigned to be 4 β -hydroxyguaioxide and 10 α -hydroxyguaioxide, respectively, providing that the configuration of the 10-position was retained.¹⁶

Oxidation of LB (14). LB (14) was subjected to the reaction with *m*CPBA (1.2 equiv) in chloroform under reflux for 12 h. The products were separated by a combination of Sephadex LH-20 and silica-gel column chromatography to isolate three compounds 15, 16, and 17, all of which were secondary alcohols (from the IR and NMR spectra, see Experimental). The first product 15 showed a molecular ion peak at *m/z* 238, suggesting the introduction of one hydroxyl group into LB (14). The proton attached to a carbon bearing the hydroxyl group appeared at δ 4.05 (m) and showed three cross peaks in its H-H COSY spectrum. The hydroxyl proton was observed at δ 3.49 as a doublet and was confirmed by the disappearance on addition of D₂O. As the NMR spectra of the compounds having a guaiane skeleton show characteristic signals of H-6 α , H-6 β , and H-7, the other protons are possible to trace starting from these protons in the COSY spectra. In this case, the H-6 α was observed at δ 1.76 (1H, d), H-6 β at δ 2.29 (1H, dd), and H-7 at δ 2.02 (1H, ddd), as the coupling constant between H-6 α and H-7 is nearly zero. The methyl group attached at C-4 position [δ 0.90 (d, *J*=7.3 Hz)] was easily distinguished by observing NOE between H-6 β . Thus both ends of the proton network of 15 were clearly assigned and other protons were traced on H-H COSY



Scheme 2. a) *m*CPBA/CHCl₃/reflux; b) Jones; c) Al₂O₃.

spectrum. Hence, the hydroxyl group was assigned at C-2 position. This conclusion was further confirmed by Jones oxidation of **15** to **18** (IR 1728 cm^{-1}) and the 2D NMR analysis of this ketone **18**. The stereochemistry was suggested by NOE difference experiments. When the proton at δ 4.05 was irradiated, NOE into 10 α -methyl group was observed. NOE's into 12- and 13-methyl groups and 4 β -proton were detected on irradiation of the hydroxyl proton at δ 3.49. Thus, the structure of **15** was established to be 2 β -hydroxy-LB.

The second product **16** showed a molecular ion peak at m/z 238 and the presence of a hydroxyl group [3450 cm^{-1} ; δ_{H} 3.73 (m); δ_{C} 78.4 (CH)]. As in the case of 2 β -ol **15**, the position of the hydroxyl group was assigned at C-3 from the H-H COSY spectrum. Since the NOE difference spectra showed NOE into the H-4 β on irradiation of the methine proton at δ 3.73 (m), the hydroxyl group at C-3 was α . The structure was further confirmed by conversion of **16** into the known enone **20**¹³⁾ through the ketone **19** [1) Jones oxidation; 2) alumina]. Thus the structure of **16** was established to be 3 α -hydroxy-LB.

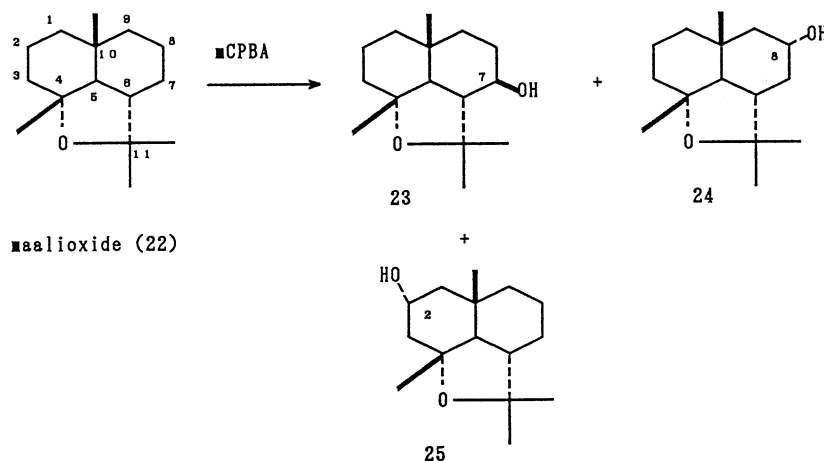
The last product **17** was also inferred as a secondary alcohol judging from the MS (m/z 238), ^1H NMR [δ 4.20 (dd)], ^{13}C NMR [δ 69.9 (CH)], and IR (3440 cm^{-1}) spectra. Although the protons at 6 α -, 6 β -, and 7 positions were characteristic, H-7 did not show the coupling with the proton attached to a carbon bearing the hydroxyl group. However, the methine proton at δ 4.20 (dd, H-8) coupled with two protons (H-9), which were further correlated to H-10. The position of hydroxyl group was suggested by oxidation of **17** into a ketone **21** (Jones oxidation), which exhibited the absorption at 1680 cm^{-1} , showing that the position of the carbonyl group was in the seven-membered ring. These results suggest that the hydroxyl group is at C-8 position. When the proton H-7 of **17** was irradiated, NOE into the 13-Me group was observed. On the other hand, when the 12-Me group was irra-

diated, NOE into H-8 was observed. These results indicate that compound **17** is 8 α -hydroxy-LB.

Oxidation of Maalioxide (22). A solution of maalioxide (**22**) in chloroform was heated at reflux with *m*CPBA for 12 h. The mixture was purified by column chromatography of Sephadex LH-20 followed by silica gel to afford alcohols **23**, **24**, and **25**, with recovered maalioxide (**22**). These three products all showed the absorption at 3400 cm^{-1} in their IR spectra and the molecular ion peak at m/z 238 in their mass spectra, suggesting that these three alcohols were all monohydroxy derivatives of maalioxide. As the ^1H and ^{13}C NMR spectra of **23** indicated the presence of a methine group [δ_{H} 3.62 (1H, td, $J=10$ and 4.4 Hz) and δ_{C} 72.8 (CH)], this alcohol must be secondary. The position of the hydroxyl group should be 7 β (equatorial), since the methine proton appeared as triplet (10 Hz) of doublets (4.4 Hz) and only this position has three protons on adjacent carbons. Hence the structure of **23** was established to be 7 β -hydroxy-maalioxide.

The second product **24** is deduced to be also a secondary alcohol from its ^1H NMR [δ_{H} 3.87 (1H, tt, $J=11$ and 3.9 Hz)] and ^{13}C NMR [δ_{C} 68.6 (CH)] spectra. As this methine proton appeared as triplet (11 Hz) of triplets (3.9 Hz), the number of protons on adjacent carbons were four and the hydroxyl group was equatorial. These observations imply that the hydroxyl group is either on 2 α or 8 α (vide infra).

From the ^1H NMR [δ_{H} 3.96 (tt, $J=11.2$ and 4.6 Hz)] and ^{13}C NMR [δ_{C} 68.1 (CH)] spectra of the last compound **25**, it was assigned to a secondary alcohol. Similar discussion on the coupling pattern of the proton attached to a carbon bearing the hydroxyl group led to the conclusion that the hydroxyl group is either on 2 α or 8 α . In order to determine the structures of **24** and **25**, either 2 α - or 8 α -hydroxymaalioxide, the NOE experiment was therefore performed. When the methine proton at δ 3.87 of **24** was irradiated, only one methyl signal appeared in the NOE difference



Scheme 3.

spectrum. On the other hand, when the proton at δ 3.96 of **25** was irradiated, NOE's into both methyl groups at C-4 and C-10 were observed. These observations clearly distinguished these two alcohols, as expected from the consideration of the model. From these results, the structures of **24** and **25** were determined to be 8 α -hydroxymaaliolide and 2 α -hydroxymaaliolide, respectively.

In the *m*CPBA oxidation reactions, the tertiary positions are easy to be oxidized giving hydroxylated compounds. In this case, however, the tertiary positions of LB (**14**) and maaliolide (**22**) were not attacked presumably because they were sterically hindered. Most of the methine protons of these compounds were β -oriented and close to the methyl groups or the epoxy bridge. The fact that all the products were equatorial alcohols was due to energetic preference and consistent with those observed before in the field of triterpenes.⁸⁾ On the contrary, guaiooxide (**2**) has many α -protons, apart from the methyl groups or epoxy bridge. Thus, the introduction of the hydroxyl groups onto guaiooxide occurred mainly from α -face. Actually 4 α -alcohol **5** was the main product and it is noteworthy that this compound **5** was also the main product of the dry ozonization¹¹⁾ or microbial oxidation,¹⁰⁾ suggesting this position was the least hindered and protruding.

Experimental

General. See Refs. 6—9.

Guaiol (1). Guaiol (**1**) was isolated from guaiac wood oil by recrystallization from acetone.

Guaiooxide (2).¹⁷⁾ A solution of guaiol (**1**) (3 g) in acetic acid (10 ml) was treated with concd H₂SO₄ (0.1 ml) at room temperature for 10 h. After extraction with ether, the extracts (2.9 g) were purified by column chromatography over silica gel (120 g) which was eluted with benzene to afford guaiooxide (**2**) (567 mg, 18.9%).

Reaction of Guaiooxide (2) with *m*CPBA. A solution of guaiooxide (**2**) (479 mg) in chloroform (50 ml) was treated with *m*CPBA (1.06 g, 2 equiv.) under reflux for 13 h. The mixture was concentrated and filtered. The filtrate was directly subjected to column chromatography on silica gel (benzene:AcOEt gradient) to give recovered guaiooxide (**2**) (230 mg), 1 β -ol **3** (6.5 mg, 2.4%¹⁸⁾), 10 α -ol **4** (8 mg, 3.0%¹⁸⁾), a mixture of **4** and **6** (ca. 5 mg), 4 α -ol **5** (26.5 mg, 9.9%¹⁸⁾). The mixture fraction was further purified by HPLC (JASCO Fine pak SIL; 10% EtOAc-hexane) to give 4 β -ol **6** (2 mg, 0.7%¹⁸⁾). 1 β -Hydroxyguaiooxide (**3**): IR (CCl₄) 3550 and 3450 cm⁻¹; ¹H NMR δ =0.93 (3H, d, *J*=6.6 Hz), 0.97 (3H, d, *J*=7.1 Hz), 1.22 (3H, s), 1.34 (3H, s), and 3.41 (1H, s, -OH); ¹³C NMR δ =22.9 (CH₃), 23.0 (CH₃), 23.1 (CH₃), 26.8 (CH₂), 30.6 (CH₂), 31.6 (CH₃), 33.2 (CH₂), 33.7 (CH₂), 35.1 (CH₂), 38.7 (CH), 46.6 (CH), 53.7 (CH), 80.7 (C), 81.6 (C), and 93.6 (C); MS 238 (M⁺), 223, 220, 205 (base), 182, and 139; HRMS: Found: *m/z* 238.1909 (M⁺). Calcd for C₁₅H₂₆O₂: M, 238.1933. 10 α -Hydroxyguaiooxide (**4**): IR (CHCl₃) 3600 and 3450 cm⁻¹; ¹H NMR δ =0.97 (3H, d, *J*=6.8 Hz), 1.15 (3H, s), 1.22 (3H, s), and 1.35 (3H, s); ¹³C NMR δ =15.7 (CH₃), 23.0 (CH₂), 23.7 (CH₃), 25.4 (CH₂), 30.4 (CH₃), 31.7 (CH₂), 34.0

(CH₃), 38.3 (CH₂), 40.4 (CH₂), 44.9 (CH), 46.0 (CH), 58.2 (CH), 73.0 (C), 82.1 (C), and 91.1 (C); MS *m/z* 238 (M⁺), 223, and 205 (base); HRMS: Found: *m/z* 238.1944 (M⁺). Calcd for C₁₅H₂₆O₂: M, 238.1933. 4 α -Hydroxyguaiooxide (**5**)^{10,11)}: IR 3600 cm⁻¹; ¹H NMR δ =0.87 (3H, d, *J*=6.1 Hz), 1.19 (3H, s), 1.308 (3H, s), and 1.315 (3H, s); ¹³C NMR δ =22.5 (CH₃), 23.0 (CH₃), 24.8 (CH₃), 24.8 (CH₂), 30.7 (CH₂), 31.1 (CH₃), 31.4 (CH₂), 33.5 (CH₂), 34.9 (CH₂), 40.0 (CH), 45.5 (CH), 52.1 (CH), 78.8 (C), 81.6 (C), and 93.0 (C); MS *m/z* 238 (M⁺), 223, 205 (base), 162, and 95. 4 β -Hydroxyguaiooxide (**6**): IR (CHCl₃) 3500 and 3400 cm⁻¹; ¹H NMR δ =0.91 (3H, d, *J*=6.6 Hz), 1.19 (3H, s), 1.22 (3H, s), and 1.32 (3H, s); ¹³C NMR δ =22.9 (CH₃), 23.0 (CH₃), 23.1 (CH₃), 26.8 (CH₂), 30.5 (CH₂), 31.6 (CH₃), 33.2 (CH₂), 33.6 (CH₂), 35.0 (CH₂), 38.7 (CH), 46.5 (CH), 53.6 (CH), 80.7 (C), 81.7 (C), and 93.6 (C); MS *m/z* 238 (M⁺), 223, 205, 162, and 95 (base); HRMS: Found: *m/z* 238.1960 (M⁺). Calcd for C₁₅H₂₆O₂: M, 238.1933.

Cyclization of Guaiol (1). A solution of guaiol (**1**) (500 mg) in acetic acid (10 ml) was treated with Pb(OAc)₄ (1.67 g, 1 equiv.) at room temperature for 24 h. Aqueous Na₂SO₃ was added and the mixture was extracted with ether. The residue was purified by column chromatography over silica gel (25 g) (benzene:EtOAc gradient) to afford a mixture of Δ^1 -dehydroguaiooxide (**7**) and Δ^1 (10)-dehydroguaiooxide (**8**) (32.7 mg).

Epoxidation of the Mixture of 7 and 8. To a solution of the mixture of **7** and **8** (86.2 mg) in chloroform (3 ml) was added *m*CPBA (144.9 mg, 1.5 equiv.). The mixture was treated with aqueous Na₂SO₃ and the residue was purified by column chromatography over silica gel (benzene:EtOAc gradient) to give 1 α ,2 α -epoxyguaiooxide (**9**) (73.6 mg), 1 β ,2 β -epoxyguaiooxide (**10**) (2.8 mg), and 1 α ,10 α -epoxyguaiooxide (**11**) (5 mg). **9**: ¹H NMR δ =0.90 (3H, d, *J*=7.6 Hz), 0.91 (3H, d, *J*=7.6 Hz), 1.16 (3H, s), 1.40 (3H, s), and 3.50 (1H, s); ¹³C NMR δ =12.8 (CH₃), 18.6 (CH₃), 23.1 (CH₃), 30.6 (CH₂), 31.1 (CH₃), 31.2 (CH₂), 34.0 (CH), 34.9 (CH₂), 35.0 (CH₂), 35.1 (CH), 44.3 (CH), 61.5 (CH), 80.8 (C), 83.0 (C), and 89.8 (C); MS *m/z* 236 (M⁺), 221, 192, 137, 55 (base), and 43; HRMS Found: *m/z* 236.1746 (M⁺). Calcd for C₁₅H₂₄O₂: M, 236.1774. **10**: ¹H NMR δ =0.73 (3H, d, *J*=7.0 Hz), 1.02 (3H, d, *J*=7.0 Hz), 1.17 (3H, s), 1.38 (3H, s), and 3.19 (1H, d, *J*=1.5 Hz); ¹³C NMR δ =17.4 (CH₃), 18.8 (CH₃), 22.8 (CH₃), 29.9 (CH₃), 30.7 (CH₂), 31.3 (CH₂), 31.9 (CH), 34.3 (CH₂), 42.1 (CH₂), 43.3 (CH), 45.1 (CH), 59.6 (CH), 68.2 (C), 82.9 (C), and 90.2 (C); MS *m/z* 236 (M⁺), 221, 203, 179, 124 (base), 109, and 43; HRMS: Found: *m/z* 236.1743 (M⁺). Calcd for C₁₅H₂₄O₂: M, 236.1774. **11**: ¹H NMR δ =1.01 (3H, d, *J*=7 Hz), 1.23 (6H, s), and 1.25 (3H, s); ¹³C NMR δ =13.3 (CH₃), 21.6 (CH₂), 27.1 (CH₃), 28.1 (CH₂), 28.4 (CH₂), 28.8 (CH₂), 28.9 (CH₃), 30.7 (CH₃), 32.5 (CH₂), 36.1 (CH), 40.9 (CH), 67.0 (C), 71.2 (C), 72.9 (C), and 75.0 (C); MS *m/z* 236 (M⁺), 221, 203, 179, 161 (base), 124, 109, and 43; HRMS Found: *m/z* 236.1743 (M⁺). Calcd for C₁₅H₂₄O₂: M, 236.1774.

Reduction of 9. A solution of **9** (30 mg) in THF (3 ml) was treated with LiAlH₄ (24 mg) under reflux for 10 h. Ether saturated with water was added and the mixture was worked up as usual to afford **12** (3 mg) after purification by column chromatography (benzene:EtOAc gradient). IR (CHCl₃) 3400 cm⁻¹; ¹H NMR δ =0.95 (3H, d, *J*=7 Hz), 1.05 (3H, d, *J*=7 Hz), 1.17 (3H, s), and 1.34 (3H, s); MS *m/z* 238 (M⁺), 223, 220, 205, and 43 (base); HRMS Found: *m/z* 238.1912 (M⁺). Calcd for C₁₅H₂₆O₂: M, 238.1933.

Reduction of 10. A solution of **10** (2 mg) in THF was

also reduced as described above to afford **3** (1.5 mg). The spectral data of this product were identical with those of the mCPBA oxidation product of guaiooxide (**2**).

Reaction of LB (14) with mCPBA. A solution of LB (**14**) (80.6 mg) in chloroform (3 ml) was treated with mCPBA (101.2 mg, 1.2 equiv.) under reflux for 12 h. The mixture was concentrated and filtered. The filtrate was directly subjected to column chromatography on Sephadex LH-20 (CHCl₃:MeOH=1:1) and then silica gel (hexane:AcOEt gradient) to give recovered LB (**14**) (46.8 mg), 2 β -ol **15** (3.3 mg, 9.1%¹⁸), 3 α -ol **16** (5.0 mg, 13.8%¹⁸), and 8 α -ol **17** (3.3 mg, 9.1%¹⁸). **15**: IR 3450 cm⁻¹; ¹H NMR δ =0.90 (3H, d, J =7.3 Hz, 1.02 (3H, d, J =6.8 Hz), 1.18 (3H, s), 1.26 (3H, s), 3.49 (1H, d, J =11.5 Hz), and 4.05 (1H, dt, J =11.5 and 5.8 Hz); ¹³C NMR δ =20.1 (CH₃), 22.1 (CH₃), 24.5 (CH₃), 25.8 (CH₂), 28.2 (CH), 31.4 (CH₃), 32.7 (CH₂), 33.4 (CH₂), 43.3 (CH), 43.4 (CH₂), 44.1 (CH), 56.9 (CH), 73.9 (CH), 84.3 (C), and 96.2 (C); MS m/z 238 (M⁺), 223, 220, and 43; HRMS Found: m/z 238.1893 (M⁺). Calcd for C₁₅H₂₆O₂: M, 238.1933. **16**: IR 3450 cm⁻¹; ¹H NMR δ =0.86 (3H, d, J =6.6 Hz), 0.87 (3H, d, J =7.6 Hz), 1.19 (3H, s), 1.25 (3H, s), and 3.73 (1H, m); ¹³C NMR δ =15.1 (CH₃), 22.2 (CH₃), 24.6 (CH₃), 25.6 (CH₂), 31.0 (CH₃), 33.5 (CH), 33.7 (CH₂), 33.9 (CH₂), 40.1 (CH₂), 43.7 (CH), 53.7 (CH), 54.6 (CH), 78.4 (CH), 83.3 (C), and 94.9 (C); MS m/z 238 (M⁺), 223, 220, and 95; HRMS Found: m/z 238.1890 (M⁺). Calcd for C₁₅H₂₆O₂: M, 238.1933. **17**: IR 3580 cm⁻¹; ¹H NMR δ =0.92 (6H, d, J =7.1 Hz), 1.24 (3H, s), 1.31 (3H, s), and 4.20 (1H, dd, J =16.4 and 6.4 Hz); ¹³C NMR δ =20.0 (CH₃), 22.4 (CH₃), 24.5 (CH₃), 28.0 (CH₂), 29.8 (CH), 30.4 (CH₂), 31.2 (CH₂), 31.8 (CH₃), 43.8 (CH₂), 44.8 (CH), 54.1 (CH), 54.6 (CH), 69.9 (CH), 81.5 (C), and 94.4 (C); MS m/z 238 (M⁺), 223, 205, and 137; HRMS Found: m/z 238.1969 (M⁺). Calcd for C₁₅H₂₆O₂: M, 238.1933.

Oxidation of 2 β -ol 15. A solution of 2 β -ol **15** (1.9 mg) in acetone (0.4 ml) was treated with Jones reagent (3 drops) at 0°C for 1 h. Usual work up and chromatography over silica gel (hexane:EtOAc gradient) gave 2-oxo-LB **18** (0.3 mg). IR 1728 cm⁻¹; ¹H NMR δ =1.09 (3H, d, J =7.3 Hz), 1.20 (3H, d, J =6.1 Hz), 1.21 (3H, s), and 2.63 (1H, dd, J =18.8 and 8.8 Hz); ¹³C NMR δ =21.1 (CH₃), 24.6 (CH₃), 25.2 (CH₃), 29.2 (CH₂), 29.7 (CH₂), 30.9 (CH₃), 34.1 (CH₂), 34.6 (CH₂), 40.0 (CH), 44.4 (CH), 45.1 (CH₂), 60.3 (CH), 83.1 (C), 92.2 (C), and 214.0 (C); MS m/z 236 (M⁺), 221, 203, and 137; HRMS Found: m/z 236.1778 (M⁺). Calcd for C₁₅H₂₄O₂: M, 236.1776.

Oxidation of 3 α -ol 16. A solution of 3 α -ol **16** (1.7 mg) in acetone (0.3 ml) was treated with Jones reagent (3 drops) at 0°C for 1 h. Usual work up and chromatography over silica gel (hexane:EtOAc gradient) gave 3-oxo-LB **19** (0.9 mg). IR 1730 cm⁻¹; ¹H NMR δ =0.91 (3H, d, J =6.7 Hz), 1.05 (3H, d, J =7.7 Hz), 1.18 (3H, s), and 1.26 (3H, s); ¹³C NMR δ =13.5 (CH₃), 22.3 (CH₃), 24.4 (CH₃), 25.3 (CH₂), 30.8 (CH₃), 33.2 (CH), 33.6 (CH₂×2), 42.1 (CH₂), 44.3 (CH), 51.7 (CH), 55.9 (CH), 83.4 (C), 90.9 (C), and 214.4 (C); MS m/z 236 (M⁺), 221, and 203; HRMS Found: m/z 236.1775 (M⁺). Calcd for C₁₅H₂₄O₂: M, 236.1776.

Isomerization of 3-Oxo LB 19.¹⁹ A solution of 3-oxo LB (**19**) (0.9 mg) in ether (1 ml) was absorbed on alumina (2.5 g) and eluted with ether (80 ml) after 5 h. Purification over silica-gel column chromatography (hexane:EtOAc gradient) afforded an enone **20** (0.2 mg). UV (MeOH) λ_{\max} =242 nm (ϵ =1.09×10⁴); IR 3400 and 1680 cm⁻¹; ¹H NMR δ =1.04 (3H, d, J =6.3 Hz), 1.23 (3H, s), 1.26 (3H, s), and 1.68

(3H, d, J =1.5 Hz); MS m/z 236 (M⁺), 218, 203, and 59.

Oxidation of 8 α -ol 17. A solution of 8 α -ol **17** (1.5 mg) in acetone (0.3 ml) was treated with Jones reagent (3 drops) at 0°C for 1 h. Usual work up and chromatography over silica gel (hexane:EtOAc gradient) gave 8-oxo-LB **21** (0.4 mg). IR 1680 cm⁻¹; ¹H NMR δ =0.96 (3H, d, J =6.6 Hz), 0.97 (3H, d, J =7.6 Hz), 1.24 (3H, s), and 1.25 (3H, s); ¹³C NMR δ =19.5 (CH₃), 22.0 (CH), 25.3 (CH₃), 28.5 (CH₂), 30.4 (CH₂), 30.7 (CH), 31.0 (CH₃), 34.4 (CH₂), 45.2 (CH), 51.6 (CH₂), 54.3 (CH), 61.1 (CH), 83.2 (C), 95.2 (C), and 202.0 (C); MS m/z 236 (M⁺), 221, 208, and 137; HRMS Found: m/z 236.1805 (M⁺). Calcd for C₁₅H₂₄O₂: M, 236.1776.

Reaction of Maalioxide (22) with mCPBA. A solution of maalioxide (**22**) (151 mg) in chloroform (10 ml) was heated at reflux with mCPBA (211 mg, 1.2 equiv.) for 12 h. The mixture was concentrated and filtered. The filtrate was directly subjected to column chromatography on Sephadex LH-20 (CHCl₃-MeOH) and then silica gel (benzene-AcOEt). Further purification by preparative TLC gave 7 β -hydroxymaalioxide (**23**) (1 mg, 1.2%¹⁸), 8 α -hydroxymaalioxide (**24**) (1.2 mg, 1.5%¹⁸), and 2 α -hydroxymaalioxide (**25**) (1.6 mg, 2.0%¹⁸) in addition to recovered maalioxide (**22**) (76 mg).

7 β -Hydroxymaalioxide (**23**); IR (CHCl₃): 3400 cm⁻¹; ¹H NMR: δ =0.92 (3H, s), 1.14 (3H, s), 1.21 (3H, s), 1.43 (3H, s), 3.62 (1H, td, J =10 and 4.4 Hz), and 3.68 (1H, s); MS m/z 238 (M⁺), 223, and 205; HRMS Found: m/z 238.1933 (M⁺). Calcd for C₁₅H₂₆O₂: M, 238.1933.

8 α -Hydroxymaalioxide (**24**); IR (CHCl₃): 3400 cm⁻¹; ¹H NMR: δ =0.92 (3H, s), 1.07 (3H, s), 1.12 (3H, s), 1.33 (3H, s), 3.67 (1H, s), and 3.87 (1H, tt, J =11 and 3.9 Hz); MS m/z 238 (M⁺), 223, 205, and 195; HRMS Found: m/z 238.1935 (M⁺). Calcd for C₁₅H₂₆O₂: M, 238.1933.

2 α -Hydroxymaalioxide (**25**); IR (CHCl₃): 3400 cm⁻¹; ¹H NMR: δ =0.93 (3H, s), 1.06 (3H, s), 1.15 (3H, s), 1.33 (3H, s), and 3.96 (1H, tt, J =11.2 and 4.6 Hz); MS m/z 238 (M⁺), 223, 205, and 147; HRMS Found: m/z 238.1931 (M⁺). Calcd for C₁₅H₂₆O₂: M, 238.1933.

We thank Prof. Takeyoshi Takahashi and Dr. Hiroshi Hirota, University of Tokyo, for their generous gift of LB and the authentic spectra. Grateful thanks are also due to Dr. Hiroshi Ishii, Shionogi Research Laboratory, Osaka, for sending the authentic spectra. This work was supported in part by a Grant-in-Aids for Cancer Research from the Ministry of Health and Welfare.

References

- 1) E.g. K. Takeda, *Pure Appl. Chem.*, **21**, 181 (1970).
- 2) E.g. Y. Asakawa, T. Ishida, M. Toyota, and T. Takemoto, *Xenobiotica*, **16**, 753 (1986).
- 3) E.g. D. H. R. Barton, J. M. Beaton, L. E. Geller, and M. M. Pechet, *J. Am. Chem. Soc.*, **82**, 2640 (1960).
- 4) E.g. R. Breslow, *Chem. Soc. Rev.*, **1**, 553 (1972).
- 5) E.g. Y. Mazur, *Pure Appl. Chem.*, **41**, 145 (1975).
- 6) Y. Asakawa, R. Matsuda, and M. Tori, *Experientia*, **42**, 201 (1986); Y. Asakawa, R. Matsuda, M. Tori, and T. Hashimoto, *Phytochemistry*, **27**, 3861 (1988).
- 7) M. Tori, R. Matsuda, and Y. Asakawa, *Bull. Chem. Soc. Jpn.*, **58**, 2523 (1985).

- 8) M. Tori, R. Matsuda, and Y. Asakawa, *Tetrahedron Lett.*, **26**, 227 (1985); M. Tori, R. Matsuda, and Y. Asakawa, *Chem. Lett.*, **1985**, 167; M. Tori, R. Matsuda, and Y. Asakawa, *Tetrahedron*, **42**, 1275 (1986).
- 9) M. Tori, M. Sono, and Y. Asakawa, *Bull. Chem. Soc. Jpn.*, **58**, 2669 (1985).
- 10) H. Ishii, T. Tozyo, M. Nakamura, and H. Minato, *Tetrahedron*, **26**, 2751 (1970).
- 11) C. -K. Ping, L. Bang, G. Ourisson, M. M. -Rohmer, E. Trifilieff, and J. -L. Zundel, *J. Chem. Res. (S)*, **1980**, 315; (*M*), **1980**, 3973.
- 12) H. Hirota, Y. Moriyama, H. Shirasaki, T. Tsuyuki, and T. Takahashi, *Bull. Chem. Soc. Jpn.*, **52**, 3755 (1979).
- 13) H. Hirota, Y. Tanahashi, and T. Takahashi, *Tetrahedron Lett.*, **1975**, 4579; *Bull. Chem. Soc. Jpn.*, **53**, 785 (1980).
- 14) T. Hashimoto, M. Tori, Z. Taira, and Y. Asakawa, *Tetrahedron Lett.*, **26**, 6473 (1985).
- 15) C. Ehret and G. Ourisson, *Bull. Soc. Chim. Fr.*, **1968**, 2629.
- 16) The stereochemistry of *m*CPBA oxidation was discussed by Müller and Schneider¹⁹⁾ and was reported to retain the original configuration. Although in our previous⁶⁾ and the present cases the products of inverted configuration at the oxidation sites were isolated, the major and minor products were those of retention and inversion of configuration, respectively.
- 17) R. B. Bates and R. C. Slagel, *Chem. Ind.*, **1962**, 1715.
- 18) Yields correspond to isolated yields based on starting material consumed.
- 19) W. Müller and H. -J. Schneider, *Angew. Chem., Int. Ed. Engl.*, **18**, 407 (1979).
-